# **TECHNICAL SHEET No. 12**

# Virus Detection: Grapevine virus A (GVA)

### Methods: RNA dot blot hybridization

#### <u>General</u>

Virus detected: GVA from grapevine leaves, petioles, and stems. General Methods are RNA dot blot hybridization.

### **Developed by**

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### Goals

To develop sensitive methods for GVA detection in grapevines.

### **Introduction**

See other technical sheets on *Grapevine virus A* for information about this virus and references.

### **Methods**

### **RNA** purification

- 1. Cut a leaf or stem grape tissue and transfer it to a microfuge tube, freeze in liquid nitrogen, and add 500  $\mu$ l 70% ethanol for 10 min.
- Discard the ethanol and add 500 μl of PEX solution; incubated at 65 °C for 5 min. (PEX solution: 6.25 mM potassium ethyl xanthogenate, 100 mM Tris- HCl pH 7.5, 700 mM NaCl, 10 mM EDTA pH 8.0).
- 3. Centrifuge the samples for 6 min at 14,000 rpm (12,000 x g) and incubate for an additional 15 min at 65 °C.
- 4. Remove the piece of tissue using a toothpick and add 1 ml ethanol and incubate for 30 min at -80 °C.
- 5. Collect nucleic acids by centrifugation for 10 min at 14,000 rpm, discard ethanol and dry gently under vacuum.
- 6. Dissolve the pellet of nucleic acid with 30  $\mu$ l ddH<sub>2</sub>O, incubate at 65 °C for 15 min and store at -80 °C.

#### **Dotting RNA on membrane**

- 1. Spot 5 µl from each RNA sample onto a nylon-membrane.
- 2. Cross-link the nucleic acid to the membrane by exposure to U.V. for 3 min.

# Preparation of <sup>32</sup>P labeled probes by Random priming method (kit from Amersham)

- 1. In a total reaction volume of 50  $\mu$ l add: 5 $\mu$ l of 0.3  $\mu$ g/ $\mu$ l plasmid pGEM/GVA, 5 $\mu$ l 10 x hexanucleotide buffer, and 22  $\mu$ l ddH<sub>2</sub>O.
- 2. Heat for 10 min at 100 °C then cool on ice for 15 min.
- 3. Add 10  $\mu$ l of dNTPs (0.5 mM each) without dCTP and 3  $\mu$ l of the Klenow fragment of DNA polymerase I.
- 4. Add 5  $\mu$ l of [<sup>32</sup>P] dCTP (3000 Ci/mmole) and incubate at 37 °C for 3 h.
- 5. Boil for 10 min and immediately put in ice bath.

### Hybridization

- 1. Membranes are prehybridized at 42 °C with a solution that contains, for 50 ml: 12.5 ml 50 x SCC, 5 ml Denhardt solution x 50, 2.5 ml 10 % SDS, 3.2 ml NaH<sub>2</sub>PO<sub>4</sub>, 1.7 ml Na<sub>2</sub>HPO<sub>4</sub>, 0.7 ml boiled salmon sperm DNA (50  $\mu$ g/ml) and ddH<sub>2</sub>O up to 50 ml.
- 2. After 2 h, add the radiolabeled probe and hybridize for 18 h at 42 °C.
- 3. Wash the blots with 1xSSC 0.1% SDS for 20 min at 65°C, twice. If background is dirty, wash with 0.1 x SSC 0.1% SDS at 65 °C.
- 4. Expose the membrane to X-ray film or to a Phosphor imager.

#### **Results**

The same samples are compared using dot blot hybridization and northern blot hybridization.

